Kinetic model of the fixation of phosphates on particles of sediments

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Abstract

The adsorption of phosphates on sediments was studied by distinguishing between the mineral and bacterial biomass compartments. Enrichment with Ca²⁺, Fe³⁺ and Al³⁺ validates a multilinear model.

Introduction

Understanding the process of the fixation of phosphorus on sediments is important for water-quality control (Overman and Chu, 1977; Boström et al., 1982). For this reason, a number of studies have been carried out on the variation of phosphorus fixed by or released from sediments with time. The kinetic equation obtained can be said to be of the first order (Fried et al., 1957; Larsen et al., 1965; Probert and Larsen, 1972) or of the second order (Kuo and Loste, 1974).

The study of the fixation of phosphates by sediments is made difficult because of the complex and diversified structure of a sediment (a mixture of living and inert matter). As a first approximation, we can consider that the sediments can be divided into two groups: one group made up of a biomass (X) and the other, a non-living group (Mi). According to this subdivision, a model can be proposed (Fig. 1):

- The biomass fixed to the sediment transfers an electrostatic potential that can, before bacterial assimilation, induce biosorption of phosphates. After biosorption, assimilation will reach a state of equilibrium, accompanied most probably by a redistribution of phosphates among the various groups.
- Other than the biomass, the abiotic groups (Fe³⁺, Ca²⁺, Al³⁺....) can also constitute sites of phosphate fixation. Nevertheless, metals are generally held as precipitated salts (carbonates...) or in fairly stable organic complexes. If the cation forms soluble salts or compounds which are not very stable, the phosphates will be able to interact strongly, probably through precipitation.

The goal of our tests was to verify the frequency with which sediments contain metals and to explain the role played by the biomass present in the kinetics of phosphate fixation and release.

Material and methods

This study of the kinetics of incorporation was carried out on sediments of freshwater lakes (Gouet, Côtes d'Armor, France). Various cations in the sediments were determined by atomic absorption. The methods used to collect and prepare the samples and conduct the analyses have been described previously (Bonzongo et al., 1989). The sediments studied present the physico-chemical characteristics as summed up in Table 1.

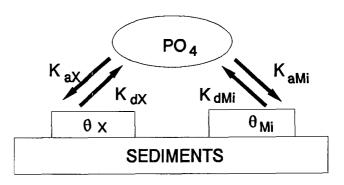


Figure 1
Model of the incorporation of phosphates in sediments

Enrichment of sediments

Multiplication of the biomass

To study the role of the microbial biomass of sediments in the fixation of phosphates, various glucose masses (0, 10, 20, 100 and 500 mg) were added to the equivalent of 5 g of dry sediments. The mixture was left to incubate at room temperature $(20 \pm 2^{\circ}\text{C})$. After bacterial growth had taken place, the following measurements were carried out: adenosine triphosphate (ATP), (Jones and Simon, 1977); the residual glucose (Dubois et al., 1965); and the phosphorus content in the different fractions according to the AFNOR T-90-023 standard, after fractionation as performed by Bonzongo et al. (1989). The equivalent of 5 g of dry sediments was then used for the kinetic incorporation.

Addition of calcium, iron and aluminium salts

Calcium, iron and aluminium were added to the sediments in the form of: CaCO₃, FeSO₄·7H₂O and Al₂ (SO₄)₃·18H₂O respectively. As in the case of glucose, the addition was accomplished by enriching the sediments separately with 0, 10, 20 and 30 mg of Ca²⁺, Fe³⁺ and Al³⁺ for 5 g of dry sediment. The sediment thus treated was used for incorporating phosphates.

Kinetic incorporation of phosphates

Sediments and the phosphate solution, prepared from KH_2PO_4 , were placed in a 100 m ℓ flask (Fig. 2) in a ratio of 1 g of dry sediment per 100 m ℓ of solution. The mixture was brought up to pH=7 using a Tris buffer and regulated by the addition of either

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TABLE 1 PHYSICO-CHEMICAL CHARACTERISTICS OF SEDIMENTS STUDIED				
Water content (% dry weight)	75.59%	Various fractions of phosphorus in sediment	P μg/g of dry sediment	
Ca ⁺⁺ µg/g of dry sediment	17400	Total Organic P	670	
Mn²+ μg/g of dry sediment	16920	Bacterial-P	179	
A1³+ μg/g of dry sediment	28000	NH ₄ CI-P	22	
MPN bacteria/g of dry sediment	109	NaOH (N/10) -P	1500	
Total P μg/g of dry sediment	2990	HCI-P	291	
		NaOH SN 85 °C	n.d.	

HC1 or NaOH. The mixtures were shaken to ensure aeration and brought up to room temperature. Sediments enriched with glucose were also aerated with an air pump. Initially the phosphorus concentration was 20 mg P/ ℓ of solution. Samples were taken with an automatic pipet to ensure homogeneity. Dissolved phosphorus was added to the samples after filtering with an 0,2 μ m sartorius filter. The experiment was performed three times and the results given correspond to the average of the three values attained.

The incorporation of the "inert group" was judged by inhibiting the biomass activity by adding 20 g of chloroform per litre of mixture.

After incorporation, the phosphates in the sediments were analysed to emphasise the sites involved in the fixation of phosphate (Bonzongo et al., 1989).

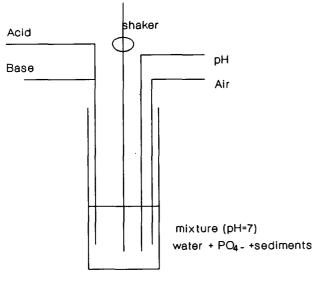


Figure 2
Experimental apparatus

Results and discussion

Nomenclature

K _{ax} and K _{aMi}	=	Constants for P-fixation
O _x	=	Mass of fixed PO ₄ per mass of biomass
O_{Mi}	=	Mass of fixed PO ₄ per mass of elements of the
		abiotic group
X	=	Biomass
Mi	=	Abiotic group
t	=	time
P_{w}	=	mg of P/e of solution
P_s	. =	mg of P/g of dry sediment
P_{sM}	=	Maximum concentration of P which can be
54.4		fixed by the sediments (mg of P/g of dry
		sediments)
P.,	=	mg of P fixed at equilibrium per unit of mass

of dry sediments

Role of the biomass

Fractionation of the phosphorus in the sediments which was performed after this treatment showed that the multiplication of the bacterial biomass of the sediments was accompanied by a transfer of the phosphorus fixed on the inert particles towards live organic groups (Fig. 3).

Residual glucose after bacterial growth had stopped was around zero for the sediments having received 10, 20 and 100 mg of glucose. However, when the added dose of glucose was 500 mg, bacterial growth stopped when approximately 50 μ g of glucose per gram of dry sediment remained. The sediments probably had a nutrient deficiency which exhausted them before all of the glucose could be consumed.

From these results, we can conclude that bacteria lacking sufficient energy and carbon for growth do use the phosphorus of the mineral group. These results confirm those of Harrison et al. (1972) which showed that isolated bacteria from Lake Klamath

could solubilise phosphates precipitated on metallic cation when a carbohydrate source of energy was present.

The kinetics of phosphate biosorption on sediment activated by glucose are represented in Fig. 4. Rapid biosorption followed by an equilibrium condition reached after 50 min can be observed. Initially it was assumed that the salts did not contain enough available metals to accomplished precipitation of the phosphate and that two sorption phenomena exist. We can then write a model of initial sorption rate (disregarding desorption):

$$\frac{dP_s}{dt} = K_{ax} Pw (\Theta_x) S + \sum K_{aMi} Pw \Theta_{Mi} S$$
 (1)

If a "doping agent" is added, it provokes an increased density of the sites of biosorption and if we assume that the desorption constants are weak, O_x increased while O_{Mi} decreases:

$$+ \frac{dP_s}{dt} = \frac{dP_{so}}{dt} + K_{ax}Pw(\Theta_x - \Theta_{xo}) + \sum K_{aMi}Pw(\Theta_{Mi} - \Theta_{Mio})$$
 (2)

If, following this activation, the inorganic contribution to fixation is "masked"; then comparing the sediment with biomass to a sterilised sediment, we obtain Fig. 5 which validates this model with:

$$\frac{1}{Pw} \left(\frac{dP_{so}}{dt} \right)_i = K_{ax} X + K_0 \tag{3}$$

It was observed that this speed corresponds with the initial biosorption. After 2 h equilibrium with assimilation was reached. The fixed phosphate will now be compared to the ATP measurement. Figure 7 shows that:

$$K_{ax} = [0,45 (ATP) + 8,3].10^{-3}$$
 $(\Delta P_{se} = 0,45 (ATP))$ (4)

We observed that during assimilation, a biomass was produced; the scale of this equilibrium can be verified thus: 1 mg of P = 0.5 g of biomass or 0.6 mg of ATP.

Role of inorganic groups

We indicated in Table 1 that the sediment which is the object of our tests contained 17 mg Fe³⁺/g dry sediment, 28 mg Al³⁺/g dry sediment, 1,7 mg Ca²⁺/g dry sediment and 2 mg Mn²⁺/g dry sediment. In our experiments, we incorporated 1 g of dry sediment /100 m² of solution, to which we added 20 mg P/ℓ. In one litre of mixture, we initially had 20 mg P for a large amount of metal (170 mg Fe³⁺, 280 mg Al³⁺, 17 mg Ca²⁺ and 20 mg Mn²⁺).

To the sediments were added Al₂SO₄.7H₂O and CaCO₃. The first hydrolised towards pH 7, the second was hardly soluble. Figures 7, 8 and 9 show the kinetics of the fixation of P by sediment enriched with Fe³⁺, Al³⁺ and Ca²⁺. They also show the importance of a weak dose of Fe³⁺ in relation to the initial iron content, thus demonstraiting the very low availability of Fe³⁺ and Al³⁺ in the sediment. The added Fe³⁺ and Al³⁺ induced a precipitation of phosphate in salt form.

Calcium carbonate, which is not very soluble, was added in the form already present in the sediment. This led to an equation similar in form to the Langmuir equation. However, a model analogous to the one present for the biomass will suffice.

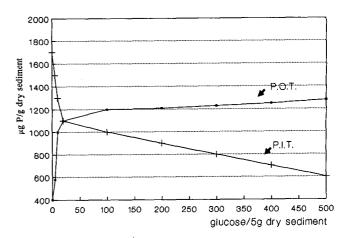


Figure 3

Evolution of the various fractions of phosphorus (TOP, TIP total and inorganic phosphorus)

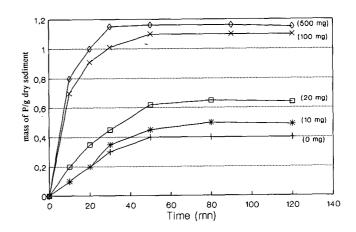


Figure 4
Kinetics of P-fixation by sediments with various biomasses
(Glucose: 0, 10, 20, 100, 500 mg)

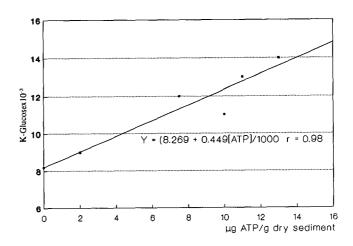


Figure 5
Fixation constants for the biomass

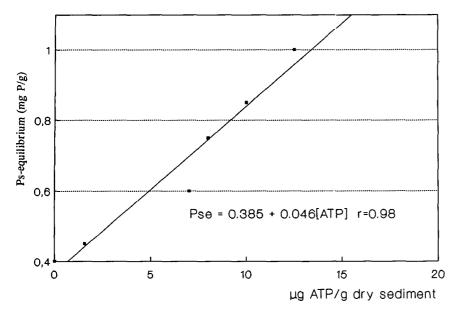


Figure 6 Relationship between phosphorus and biomass at equilibrium

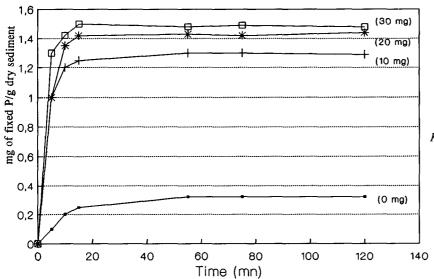


Figure 7
Kinetics of P-fixation by sediment enriched with iron (0, 10, 20, 30 mg Fe+++/5g dry sediment)

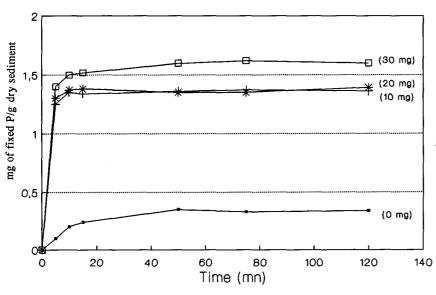


Figure 8
Kinetics of P-fixation by sediment enriched with Aluminium (0, 10, 20, 30 mg Al***/5g dry sediment)

1,2 1 (30 mg) E 1 (20 mg) * (10 mg) + (0 mg) (0 mg) (0 mg) (0 mg)

Figure 9
Kinetics of P-fixation by sediment enriched with calcium (0, 10, 20, 30 mg Ca++/5g dry sediment)

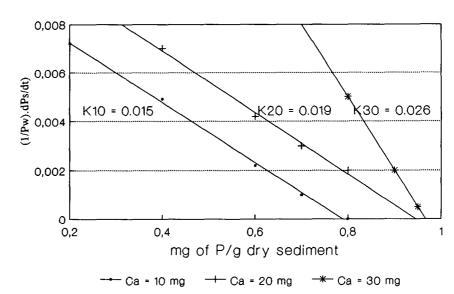


Figure 10
Constants for P-fixation by sediments with various calcium concentrations

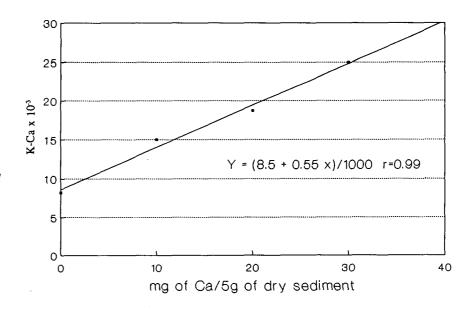


Figure 11
Relationship between the fixation
constants and the mass of calcium added

$$\frac{1}{P_{\cdots}} \left(\frac{dP_s}{dt} \right) = K_{aMi} \left(O_M \right) \cdot S = K_{ca} \left(P_{sM} - P_s \right) \tag{5}$$

We found that the curve is positive with respect to the mass added (Fig. 11). $K_{CaCO3} = (8.5 + 0.552.Ca)10^{-3}$ with Ca^{2+} corresponding to the mass of calcium per mass of dry sediment.

 K_{CaCO3} increases linearly with the mass of $CaCO_3$ corresponding to the increase of the surface transfer of phosphates to carbonate. Therefore P_{sM} also seems to increase with the level of incorporation of soluble salt.

To summarise:

- The Fe³⁺ and the Al³⁺ in the sediments studied are not readily available for the fixation of phosphate and the incorporation of a relatively low amount of available Fe³⁺ or Al³⁺ induces a large elimination of phosphate by chemical "dephosphorisation".
- Incorporation of Ca²⁺ in the sediments with amounts identical to those of Fe³⁺ and Al³⁺ favors only a weak elimination of phosphate.

Conclusion

Incorporation of phosphates in sediments is dependent upon the composition of the materials. In our experiments, we have shown that a model of biosorption could be applicable if the sediments containing the bacteria and the phosphatic liquor are well mixed. After a fairly short time, the phosphates that assimilated well correlated with the biomass of the materials.

As for the fixation of phosphates in abiotic groups, it seems that the metals Fe³⁺, A1³⁺ and Ca²⁺ play an important role, but that the availability of metals for the incorporation of phosphates in

sediments is very low, so a very small dose of soluble salts induces a large elimination, which is not observed in non-treated material. Precipitated salts which are not very soluble, or form stable compounds, create sites of physio-sorption with weak affinity.

References

- BONZONGO, JC, BERTRU, G and MARTIN, G (1989) Les méthodes de spéciations du phosphore dans les sédiments: critiques et propositions pour l'évaluation des fractions minérales et organiques. *Arch. Hydrobiol.* **116**(1) 61-69.
- BOSTRÖM, B, JANSON, M and FORSBERG, C (1982) Phosphorus release from lake sediments. *Arch. Hydrobiol.* Beich Ergeb. Limnol. **18** 5-59.
- DUBOIS, M, GILLES, KA, HAMILTON, JK, RIBERS, PA and SMITH, F (1956) Colorimetric method for determination of sugars and related substances. *Analytical Chem.* **28** 350-356.
- FRIED, M, HAGEN, CE, SAIZ, DELRIO and LEGGERT, JE (1957) Kinetics of phosphate uptake in the soil plant system. J. Soil Sci. 84 427-437.
- HARRISON, MJ, PACHA, RE and MORITA, RY (1972) Solubilization of inorganic phosphorus by bacteria isolated from upper Klamath lake sediment. *Limnol. Oceanogr.* 17 50-57.
- JONES, JG and SIMON, BM (1977) Increased sensivity in the measurement of ATP in freshwater samples with a comment on the adverse effect of membrane filtration. Freshwater Biology 7 253-260
- KUO, S and LOSTE, EG (1974) Kinetics of phosphate adsorption and desorption by hematite and gibbsite. J. Soil Sci. 116 400-406.
- LARSEN, S, GUNARY, D and SUTTON, CD (1965) The rate of immobilization of applied phosphate in relation to soil properties. J. Soil Sci. 16 141-148.
- PROBERT, ME and LARSEN, S (1972) The kinetics of heterogeneous isotopic exchange. J. Soil Sci. 23 76-81.
- OVERMAN, AR and CHU, RL (1977) A kinetic model of steady state phosphorus in a batch reactor. I. Effect of soil/solution ratio. Water Research 11 771-775.