

# Optimisation of the PCR-*invA* primers for the detection of *Salmonella* in drinking and surface waters following a pre-cultivation step

**KLM Moganedi<sup>1\*</sup>, EMA Goyvaerts<sup>2</sup>, SN Venter<sup>3</sup> and MM Sibara<sup>4</sup>**

<sup>1</sup>Department of Microbiology, University of Limpopo, Private Bag X1106, Sovenga, 0727, South Africa

<sup>2</sup>Kitso Biotech Pty Ltd, 23 Fernvillia P1, Pietermaritzburg 3201, South Africa

<sup>3</sup>Department of Microbiology and Plant Pathology, University of Pretoria, Pretoria 0002, South Africa

<sup>4</sup>Department of Education, Private Bag X603, Pretoria 0001, South Africa

## Abstract

A polymerase chain reaction (PCR)-based method for the detection of *Salmonella* species in water samples was optimised and evaluated for speed, specificity and sensitivity. Optimisation of Mg<sup>2+</sup> and primer concentrations and cycling parameters increased the sensitivity and limit of detection of PCR to 2.6 x 10<sup>4</sup> cfu/ml. A 6h non-selective pre-enrichment step further increased the limit of detection to 26 cfu/ml. Out of 14 different *Salmonella* strains tested, only two, *Salmonella arizonae* and *Salmonella pullorum*, did not give positive amplification results with primers homologous to a conserved region of the *invA* gene. When environmental and drinking waters were assessed, a non-selective pre-enrichment step was included to increase the detection efficiency of PCR. The PCR method demonstrated specificity in the presence of other competing micro-organisms as confirmed by the conventional culture method. No false positives or negatives were observed when household and environmental water samples were tested by *invA*-PCR analysis parallel to the culture method.

**Keywords:** water quality, *Salmonella*, PCR, *invA* primers